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OPFRs in urine

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1 Summary

Within the frame of the HBM4EU project, an External Quality Assurance Scheme (EQUAS) was organised on the determination of four OPFR biomarkers in urine. This was the 4th ICI/EQUAS round for this substance group within the HBM4EU program.

In total, 14 laboratories were invited for this 4th ICI/EQUAS and six laboratories (including three expert laboratories) submitted results. Four labs were able to report results for all four parameters and two labs reported results for three parameters.

In December 2019, each participant received one tube of burdened control materials of human urine (low level – level 1), one tube of burdened control materials of human urine (high level – level 2). The biomarker concentrations were approximately in the range of 2-6 µg/L and 8-25 µg/L for level 1 and level 2, respectively. The concentrations were chosen according to the review of relevant data on the occurrence of OPFRs in urine of the European population published mostly during the last five years.

A homogeneity assessment showed that both materials were sufficiently homogeneous for EQUAS testing. No issues with stability of testing materials occurred for OPFRs.

The determination of expert value based on results from expert laboratories was not possible only for BCEP, because the minimum number of expert results wasn't reached for this biomarker.

The evaluation of laboratory performance using Z-scores was performed for DPHP, BDCIPP and BCIPP for the first time. Most laboratories showed well comparable results for BCEP too, although a regular evaluation was not possible. The achieved results for BCEP are present in the report for further comparison between participants and expert labs.

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2 Introduction

Interlaboratory Comparison Investigations (ICI) and External Quality Assurance Schemes (EQUAS) are tools to assess the proficiency of laboratories, and the comparability and reliability of analytical methods. Participation in ICI/EQUAS forms an integral part of quality control, in addition to initial and on-going in-house method validation.

This 4th ICI/EQUAS study has been organised within the frame of HBM4EU as part of the Quality Assurance program for biomonitoring analyses, following protocols HBM4EU-SOP-QA-001 to 004 which are available through the HBM4EU website (<https://www.hbm4eu.eu/online-library/>). Within HBM4EU, participation in ICI/EQUAS exercises is mandatory for laboratories that will analyse HBM4EU samples.

This report describes the 4th ICI/EQUAS for OPFRs in urine, which was conducted as EQUAS and was organised by UCT Prague (University of Chemistry and Technology, Prague; VŠCHT, Vysoká škola chemicko-technologická v Praze), Department of Food Analysis and Nutrition. The analyses for homogeneity and stability testing were performed by the partner laboratory IPASUM (Institut und Poliklinik für Arbeits-, Sozial- und Umweltmedizin der Universität Erlangen-Nürnberg).

For this 4th ICI/EQUAS, expert laboratories had to be selected according to the selection criteria described in HBM4EU-SOP-QA-001 and in agreement with the QAU.

The selection of the most relevant OPFRs was previously done in WP9, and has been described in Deliverable report 9.2 v1.1. Based on this, a set of four target biomarkers was compiled to be included in the EQUAS for OPFR analysis in urine.

EQUAS is similar to ICI but instead of using the consensus value as assigned value, the mean concentration as established from data generated by at least three designated expert laboratories is used. As in an ICI, Z-scores are calculated as a measure of proficiency.

2.1 Confidentiality

In this report the identity of the participants and the information provided by them are treated as confidential. However, lab codes of the participants will be disclosed to the HBM-QAU for performance assessments.

3 Control material

3.1 Preparation of control material

The human urine was collected from one person during one day. A total of two litres were obtained. Urine was placed into the refrigerator at 7 °C overnight. The next day the sediment was centrifuged and filtrated. The whole procedure was repeated twice.

Before the spiking procedure, the urine was thawed at room temperature (20 °C). Then it was stirred for 30 min in a 3 L beaker using a magnetic stirrer. After that, three aliquots (500 mL in graduated cylinder) were transferred into the 1 L beaker (one aliquot for urine level 1 and one for urine level 2). Individual OPFR delivered as solids were dissolved with respect to the manufacturers' recommendations. Subsequently, each standard of the biomarker was appropriately diluted into methanol and individually spiked into the urine level 1 and urine level 2 using calibrated Eppendorf Multipette®. During the spiking procedure, the urine was mixed using a magnetic stirrer for the whole time, and when all compounds had been added, subsequent mixing for 30 minutes was performed. A total of 10 mL from "blank" urine, level 1 and level 2 urine was placed into the tube and later analysed for homogeneity testing. For the ICI/EQUAS and stability testing, a total of 5 mL was placed into the tube from each prepared material (urine level 1, urine level 2). All tubes were placed into the freezer at -18 °C before analysis / dispatch.

3.2 Homogeneity of control material

The homogeneity of the control material was tested according to HBM4EU-QA-002. Ten tubes of control material at level 1 and level 2 were randomly selected from the freezer and sent to IPASUM for analysis. The GC-MS/MS-based method for the detection of OPFR metabolites in human urine after solid phase extraction and derivatization with pentafluorobenzylbromide was used (Fromme et al. 2014).

The mean concentrations and relative standard deviations (RSDr) as obtained during a homogeneity testing are presented in **Table 1**. The statistical evaluation of level 1 and level 2 materials for each of the biomarkers is provided in **Appendix 1**. It was concluded that homogeneity was adequate for all quantified biomarkers at both levels.

Table 1: Concentration of OPFRs as obtained during homogeneity testing (for details see Appendix 1)

Biomarker	Level 1 (low)		Level 2 (high)	
	Mean (µg/L)	RSDr (%)	Mean (µg/L)	RSDr (%)
BCPP	5.915	3	29.342	4
BCEP	3.448	4	12.474	3
DHPH	2.426	7	8.484	5
BDCPP	4.833	10	16.262	8

3.3 Stability of control material

The stability of the control material was tested according to HBM4EU-QA-002. On the day of preparation of the control materials, randomly selected test urine samples of level 1 and level 2 were stored at -80 °C. After the deadline of submission of analysis results by the participants six test samples of both materials stored at -80 °C and six samples of both materials randomly selected from

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the -18 °C freezer, where the ICI samples were stored, were selected for analysis by IPASUM. For the analysis the previously described methods were used (see 3.2 Homogeneity of control material). The stability was evaluated using the Excel-sheet “HBM4EU ICI-EQUAS stability test CM v1”. The results are presented in **Appendix 2**. In summary, no troubles with the stability were identified.

4 Organisational details

4.1 Participants

For the organisation of the 4th ICI/EQUAS, IPASUM conducted a survey to find expert laboratories for the analysis of OPFRs in urine willing to participate in the project. Then, IPASUM evaluated their eligibility and selected expert laboratories in agreement with the QAU and according to HBM4EU-SOP-QA-001.

UCT Prague contacted the selected expert laboratories and sent them invitation letters by e-mail. It was indicated that participation would be free of charge, and that those who subscribed to this EQUAS would receive a kit containing the test materials needed for analysis. The final number of expert labs was three, all of them from the HBM4EU consortium.

Participants of this 4th ICI/EQUAS were laboratories from the HBM4EU consortium (including linked-third parties) that had been included as candidate laboratories for analyses in the frame of the HBM4EU project through WP9 (Task 9.2, Deliverable 9.3). Invitation letters (**Appendix 3**) and registration forms (**Appendix 4**) were sent by e-mail on 14/10/2019 to 14 laboratories. For registration, each participant was asked to provide which of four biomarkers were included in their scope. The participants were informed that the participation will be free of charge. The deadline for registration was 10/11/2019. Out of 14 invited laboratories, six labs (including three expert labs) agreed to participate. All registered laboratories submitted results.

4.2 Dispatch and instructions

Test materials were dispatched on 02/12/2019. Each participant received one tube of burdened control materials of human urine (low level – level 1) and one tube of burdened control materials of human urine (high level – level 2). Each sample consisted of approximately 5 mL urine.

Moreover, a letter with instructions on sample handling (**Appendix 5**), a sample receipt form to be sent back to UCT Prague upon receipt of the test material (**Appendix 6**) as well as a result submission form and a method information form (**Appendix 7**) were sent to the participants by e-mail. The latter form was used to extract relevant information related to the analytical method used for quantification.

Test materials were dispatched to the expert laboratories under frozen conditions (on dry ice) on 02/12/2019. Each lab received six tubes of burdened control materials of urine (low level – level 1) and six tubes of burdened control materials of urine (high level – level 2). Each sample consisted of approximately 5 mL urine.

Participants and expert labs were asked to perform a single analysis of each sample using the same procedure as will be used for analysis of samples in the frame of HBM4EU and to report results following the instructions given. The deadline for submitting results was 20/01/2020.

4.3 Deviations from ICI/EQUAS SOPs

For this 4th ICI/EQUAS, the HBM4EU-QA-SOPs (version 2) were followed. There were no deviations from these SOPs.

5 Data evaluation

5.1 False positives and <LOQ

Classification of false positives and biomarkers reported as "<LOQ-value" or "not detected" is done as described in HBM4EU-SOP-QA-003.

A result was assigned as false positive when all of the following conditions applied:

- 1) the biomarker is below the LOQ value as applied by the organiser, the expert laboratories, and the majority of the participants.
- 2) the biomarker is reported by the participant at a level clearly exceeding the LOQs mentioned under 1).

When a biomarker is reported as "<LOQ-value", AND an assigned value could be established for the biomarker in the control material, a further assessment was done to verify whether this result might be a false negative and to judge whether the LOQ is considered adequate (low enough) for analysis in the frame of HBM4EU. A result is a false negative when the LOQ of a biomarker is well below the assigned value, but the laboratory did not report a quantitative value. The LOQ is considered not adequate (too high) when:

- 1) the LOQ is substantially above the assigned value
- 2) the assigned value represents a realistic concentration of real samples in the frame of HBM4EU
- 3) quantitative determination is feasible by the majority of laboratories

In order to judge "<LOQ" results in a quantitative way, 'proxy-Z-scores' are calculated as described in 5.6.

5.2 Assigned value

For EQUAS studies, the concentration as established by expert laboratories is used as assigned value. The expert-assigned value is the target value based on analysis results obtained from analysis of the control material by at least three expert laboratories (see HBM4EU-SOP-QA-001). In brief, using the individual means of the expert laboratories, the mean-of-means was calculated and its relative uncertainty. The mean of means is used as assigned value when the relative uncertainty was below $0.7 \cdot \sigma_T$. If this condition is not met, and no outliers could be identified, then the uncertainty of the expert-derived mean is considered too high to be used as assigned value. The other requirement to be met is that the number of (remaining) individual expert means had to be at least three.

In case no expert value could be obtained, the consensus value derived from the combined results from both participants and expert laboratories is used as an alternative, but this is subject to a minimum of seven results in total. In this case the consensus value is calculated using robust statistics as described for ICI in HBM4EU-SOP-QA-003.

5.3 Target standard deviation (σ_T)

For calculation of the Z-scores, a fit-for-purpose relative target standard deviation (FFP-RSDR) of 25% of the assigned value is used as target standard deviation. This was the default indicated in HBM4EU-SOP-QA-003 and considered appropriate based on the outcome of the 1st, 2nd and 3rd rounds.

5.4 Z-scores

Z-scores are calculated according to SOP HBM4EU-SOP-QA-003.

$$Z = \frac{x - C}{\sigma_T} \quad (1)$$

with: Z = Z-score for the submitted analysis result;
 x = result submitted by the laboratory;
 C = expert-assigned value;
 σ_T = target standard deviation, here 0.25*C

In accordance with ISO 13528 and ISO 17043 and the deliverable D 9.4 “*The Quality Assurance/Quality Control Scheme in the HBM4EU project*”, Z-scores are classified as presented in **Table 2**.

Table 2: Classification of Z-scores

$ Z \leq 2$	Satisfactory
$2 < Z < 3$	Questionable
$ Z \geq 3$	Unsatisfactory

5.5 Proxy-Z-scores

'Proxy-Z-scores' are used here to judge "<LOQ" results in a quantitative way (see 5.1). The proxy-Z-scores' are calculated using the LOQ-value as result and equation (1). If no LOQ is specified, zero is used.

Proxy-Z-scores are classified as follows:

proxy-Z \leq -3	false negative. Based on the LOQ provided, the laboratory should have been able to detect and quantify the biomarker. Performance is considered 'unsatisfactory'.
proxy-Z \geq 3	the LOQ is considered too high to be fit-for-purpose in the frame of HBM4EU analysis. It also means that the LOQ is too high in comparison with other laboratories. (Note: proxy-Z can only be calculated when an assigned value could be established. If this is the case, this inherently means that reliable quantitative determination at a certain low level is feasible). Performance is considered 'unsatisfactory'.
-3 \leq proxy-Z < -2	possible false negative. Performance is considered 'questionable'.
2 < proxy-Z \leq 3	the LOQ is relatively high in relation to HBM4EU analysis and compared to other laboratories. Performance is considered 'questionable'.
-2 \leq proxy-Z \leq 2	LOQ is within an acceptable range relative to the assigned value, adequate for HBM4EU analysis, and in line with the LOQs of the majority of the participating laboratories. Performance is considered 'satisfactory'.

6 Results and discussion

6.1 Results submitted by participants

In total, six laboratories including three expert labs agreed to participate in this study and all of them submitted results. Two expert labs reported six results for each analysed urine sample. As described above, the urine material was sent to IPASUM for homogeneity testing. This lab is also involved as an expert in this round.

Four labs were able to report results for all four parameters (DHP, BDCIPP, BCIPP, BCP) and two labs reported results for three parameters (DHP, BDCIPP, BCIPP). The provided LOQs were comparable between participants (**Table 3**).

Table 3: Scope and LOQs ($\mu\text{g/L}$) as provided in the method information submitted by the laboratories

Lab code	DHP	BDCIPP	BCP	BCIPP	Total
PT4OPFR01 (expert)	0.05	0.02	NA	1	3
PT4OPFR03	0.03	0.09	0.3	0.3	4
PT4OPFR04 (expert)	0.15	0.1	0.1	0.2	4
PT4OPFR05 (expert)	0.1	0.5	NA	6	3
PT4OPFR06	0.1	1.3	2.3	0.2	4
PT4OPFR07	0.3	0.02	0.5	0.2	4
Total	6	6	4	6	

Table 4 gives an overview of all results reported by both expert and candidate laboratories. Regarding BCIPP at level 2, five out of six labs provided comparable results. In the case of BCP at level 2, three out of four labs reported similar results.

Table 4: The comparison of results reported by participating laboratories

	DHP level 1	DHP level 2	BDCIPP level 1	BDCIPP level 2	BCP level 1	BCP level 2	BCP level 1	BCP level 2
No. of candidates + experts	3+3	3+3	3+3	3+3	3+3	3+3	3+1	3+1
No. of quantitative results	6	6	6	6	6	6	4	4
Study RSD _R for all results (%)	17	11	12	11	17	26	16	29
Lab code	Results ($\mu\text{g/L}$)							
PT4OPFR01 (expert)	2.295	8.089	4.386	13.592	5.668	23.673	NA	NA
PT4OPFR03	2.370	8.600	3.950	12.000	5.350	24.300	3.290	11.640
PT4OPFR04 (expert)	2.468	8.454	5.170	16.993	5.903	29.851	3.448	12.474
PT4OPFR05 (expert)	2.552	8.864	4.434	14.180	4.864	26.671	NA	NA
PT4OPFR06	3.572	11.074	5.763	14.636	3.344	10.733	4.140	9.253
PT4OPFR07	2.346	8.254	4.754	15.759	4.912	21.840	4.912	19.644

6.2 Assigned values and (target) standard deviations

Using the individual means of the expert values, the mean-of-means was calculated and its relative uncertainty. The mean-of-means can be used as assigned value if the relative uncertainty is below $0.7 \cdot \sigma_T$. This condition was met for DPHP, BDCIPP and BCIPP at both levels.

Calculation of expert value for BCEP was not possible because of the small number of experts' results (minimum is three expert labs). Alternative approach based on calculation of the consensus value derived from the combined results from both participants and expert laboratories was also not possible, because the number of labs' results needed for the robust statistic was smaller than seven.

6.3 Assessment of laboratory performance

The assessment of laboratory performance was possible for most biomarkers and it is summarized in **Table 5**, **Table 6** and **Table 7**. A graphical presentation of the Z-scores is provided in **Appendix 8**. The number of satisfactory scores ($-2 < \text{Z-score} < 2$) was 100 % for DPHP and BDCIPP at both levels. Regarding BCIPP the number of satisfactory scores was 100 % at level 1 and 83 % at level 2.

Table 5: Assigned values and participant's performance for DPHP

	DPHP: Level 1			DPHP: Level 2		
Z-score based on	expert value ($n_{\text{expert labs}}=3$)			expert value ($n_{\text{expert labs}}=3$)		
No. of participants	6			6		
No. of quantitative results	6			6		
Expert value (ng/ml)	2.438			8.469		
Uncertainty of assigned value (ng/ml)	0.062			0.183		
Relative uncertainty (%)	2.5			2.2		
Relative FFP-target standard deviation (%)	25			25		
Study RSDr (%)	19			12		
	Value	Z-score	Classification	Value	Z-score	Classification
PT4OPFR01	2.295	-0.24	satisfactory	8.089	-0.18	satisfactory
PT4OPFR03	2.370	-0.11	satisfactory	8.600	0.06	satisfactory
PT4OPFR04	2.468	0.05	satisfactory	8.454	-0.01	satisfactory
PT4OPFR05	2.552	0.19	satisfactory	8.864	0.19	satisfactory
PT4OPFR06	3.572	1.86	satisfactory	11.074	1.23	satisfactory
PT4OPFR07	2.346	-0.15	satisfactory	8.254	-0.10	satisfactory

Table 6: Assigned values and participant's performance for BDCIPP

	BDCIPP: Level 1			BDCIPP: Level 2		
Z-score based on	expert value (n _{expert labs} =3)			expert value (n _{expert labs} =3)		
No. of participants	6			6		
No. of quantitative results	6			6		
Expert value (ng/ml)	4.663			14.922		
Uncertainty of assigned value (ng/ml)	0.207			0.857		
Relative uncertainty (%)	4.4			5.7		
Relative FFP-target standard deviation (%)	25			25		
Study RSDr (%)	14			12		
	Value	Z-score	Classification	Value	Z-score	Classification
PT4OPFR01	4.386	-0.24	satisfactory	13.592	-0.36	satisfactory
PT4OPFR03	3.950	-0.61	satisfactory	12.000	-0.78	satisfactory
PT4OPFR04	5.170	0.43	satisfactory	16.993	0.56	satisfactory
PT4OPFR05	4.434	-0.20	satisfactory	14.180	-0.20	satisfactory
PT4OPFR06	5.763	0.94	satisfactory	14.636	-0.08	satisfactory
PT4OPFR07	4.754	0.08	satisfactory	15.759	0.22	satisfactory

Table 7: Assigned values and participant's performance for BCIPP

	BCIPP: Level 1			BCIPP: Level 2		
Z-score based on	expert value (n _{expert labs} =3)			expert value (n _{expert labs} =3)		
No. of participants	6			6		
No. of quantitative results	6			6		
Expert value (ng/ml)	5.478			26.732		
Uncertainty of assigned value (ng/ml)	0.257			1.456		
Relative uncertainty (%)	4.7			5.4		
Relative FFP-target standard deviation (%)	25			25		
Study RSDr (%)	18			29		
	Value	Z-score	Classification	Value	Z-score	Classification
PT4OPFR01	5.668	0.14	satisfactory	23.673	-0.46	satisfactory
PT4OPFR03	5.350	-0.09	satisfactory	24.300	-0.36	satisfactory
PT4OPFR04	5.903	0.31	satisfactory	29.851	0.47	satisfactory
PT4OPFR05	4.864	-0.45	satisfactory	26.671	-0.01	satisfactory
PT4OPFR06	3.344	-1.56	satisfactory	10.733	-2.39	questionable
PT4OPFR07	4.912	-0.41	satisfactory	21.840	-0.73	satisfactory

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6.4 Conclusions and recommendations

In this HBM4EU 4th ICI/EQUAS on OPFR biomarkers in urine, 14 laboratories were invited, of which six submitted results. The overall participation rate was 43%. Two test materials were provided to each participant (spiked material at low level 1 and spiked material at high level 2).

Quantitative performance using Z-scores was assessed for DPHP, BCIPP and BDCIPP at both levels. The number of satisfactory scores ($-2 < Z\text{-score} < 2$) was 100 % for DPHP and BDCIPP at both levels. Regarding BCIPP the number of satisfactory scores was 100 % at level 1 and 83 % at level 2.

As explained above, the determination of assigned value was not possible for BCEP, for which the number of expert labs was smaller than three and there was no possibility to calculate a consensus value due to the small number of participants. Nevertheless, most labs showed well comparable results.

In conclusion, the calculation of the assigned value for three biomarkers (DPHP, BDCIPP and BCIPP) and Z-score determination was realized for the first time. As recognized in the previous three ICI/EQUAS, the determination of OPFR metabolites in urine is very challenging. For this reason a Webex meeting had been organized by UCT Prague and IPASUM after the 3rd Round. The discussion with candidate labs proposed several steps towards the harmonization of the analytical method which can increase the result quality. This 4th ICI/EQUAS documents great improvement of participants' performance for three OPFR biomarkers. Finally, a quite significant core network of satisfactory laboratories is qualified for the analysis of real samples.

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7 References

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Appendix 1: Homogeneity data

	BCPP - level 1		BCEP - level 1		DPP - level 1		BDCPP - level 1	
	Replicate 1	Replicate 2	Replicate 1	Replicate 2	Replicate 1	Replicate 2	Replicate 1	Replicate 2
1	5.798	5.863	3.478	3.315	2.605	2.622	5.230	5.380
2	5.915	5.966	3.493	3.477	2.573	2.381	4.870	5.380
3	5.870	5.979	3.538	3.533	2.467	2.730	5.340	5.300
4	5.825	5.911	3.410	3.393	2.474	2.322	4.930	4.260
5	6.195	6.207	3.334	3.500	2.446	2.454	4.280	4.280
6	6.086	6.244	3.567	3.372	2.568	2.484	4.740	5.050
7	5.930	5.947	3.297	3.230	2.348	2.279	5.290	4.470
8	5.811	5.890	3.658	3.674	2.117	2.378	4.560	4.680
9	5.711	5.686	3.278	3.613	2.511	2.083	4.420	4.370
10	5.616	5.844	3.352	3.438	2.238	2.441	4.680	4.440
Grand mean	5.915		3.448		2.426		4.798	
Cochran's test								
C	0.4701		0.5158		0.4220		0.4266	
C crit	0.8674		0.8674		0.8674		0.8674	
C < Ccrit ?	no outliers detected		no outliers detected		no outliers detected		no outliers detected	
σ_T	1.4787		0.8619		0.6065		1.1994	
s_x	0.1634		0.1052		0.1281		0.3671	
s_w	0.0743		0.1041		0.1473		0.2807	
s_s	0.1547		0.0751		0.0745		0.3088	
c crit	0.3904		0.2275		0.0998		0.3166	
Ss < c	homogeneity adequate		homogeneity adequate		homogeneity adequate		homogeneity adequate	
sw < 0.5 σ_T ?	method suited		method suited		method suited		method suited	
	BCPP - level 2		BCEP - level 2		DPP - level 2		BDCPP - level 2	
	Replicate 1	Replicate 2	Replicate 1	Replicate 2	Replicate 1	Replicate 2	Replicate 1	Replicate 2
1	29.783	30.100	12.124	12.486	8.564	8.290	16.050	15.140
2	29.349	30.391	12.595	12.266	8.471	8.187	17.640	17.750
3	30.022	29.463	12.438	12.548	8.277	8.935	17.780	17.600
4	29.907	30.098	12.503	12.988	9.067	8.477	16.130	17.700
5	29.957	29.192	12.449	12.801	8.722	8.969	16.060	16.210
6	29.928	29.073	12.459	12.376	8.251	8.461	17.430	17.100
7	29.814	30.463	12.491	12.313	7.849	8.395	16.590	17.300
8	28.152	28.930	11.961	12.476	7.669	8.944	14.000	15.040
9	27.307	29.215	12.279	12.766	8.233	9.156	14.230	15.510
10	26.179	29.522	11.753	13.414	7.866	8.889	15.030	15.940
Grand mean	29.342		12.474		8.484		16.312	
Cochran's test								
C	0.5979		0.7056		0.3342		0.3277	
C crit	0.8674		0.8674		0.8674		0.8674	
C < Ccrit ?	no outliers detected		no outliers detected		no outliers detected		no outliers detected	
σ_T	7.3356		3.1186		2.1209		4.0779	
s_x	0.8153		0.1548		0.2335		1.1549	
s_w	0.9668		0.4422		0.4932		0.6132	
s_s	0.4444		0.2717		0.2591		1.0704	
c crit	1.9366		0.8233		0.5599		1.0766	
Ss < c	homogeneity adequate		homogeneity adequate		homogeneity adequate		homogeneity adequate	
sw < 0.5 σ_T ?	method suited		method suited		method suited		method suited	

Appendix 2: Stability data

Biomarker	BCPP level 1		BCPP level 2		BCEP level 1		BCEP level 2	
time (days)	0	40	0	40	0	40	0	40
	5.947	6.220	30.100	29.510	3.478	3.530	12.124	11.890
	6.195	6.070	30.022	31.080	3.315	3.540	11.266	11.580
	6.207	5.870	30.098	29.420	3.297	3.320	12.376	11.750
	6.086	5.950	29.192	29.360	3.230	3.260	11.961	11.740
	6.244	6.280	29.073	30.680	3.493	3.250	12.279	11.830
	5.890	5.860	30.463	30.430	3.477	3.350	11.753	12.170
Average	6.095	6.042	29.825	30.080	3.382	3.375	11.960	11.827
Std dev	0.147	0.179	0.559	0.743	0.115	0.129	0.407	0.198
x0-xa (difference)	0.053		-0.255		0.007		0.133	
Test 'consequential instability':								
σ_H	1.34		6.56		0.74		2.63	
$0,3*\sigma_H$	0.40		1.97		0.22		0.79	
x0-xa<0,3* σ_H ? (consequential instability)	NO		NO		NO		NO	
Test 'significant difference':								
t	0.56		0.67		0.09		0.72	
t-crit	2.23		2.23		2.23		2.23	
Significant difference	NO		NO		NO		NO	
Biomarker	BDCPP level 1		BDCPP level 2		DPP level 1		DPP level 2	
time (days)	0	40	0	40	0	40	0	40
	4.280	5.080	15.140	16.080	2.605	2.440	8.564	7.840
	4.260	4.590	16.130	15.740	2.622	2.520	8.290	8.550
	4.560	4.730	17.700	15.550	2.348	2.530	8.471	8.950
	4.680	4.440	16.590	15.590	2.279	2.560	8.187	8.220
	4.420	4.660	15.040	15.140	2.573	2.830	8.277	7.870
	4.370	4.560	14.230	15.860	2.381	2.560	8.935	9.160
Average	4.428	4.677	15.805	15.660	2.468	2.573	8.454	8.432
Std dev	0.164	0.220	1.251	0.319	0.149	0.133	0.273	0.552
x0-xa (difference)	-0.248		0.145		-0.105		0.022	
Test 'consequential instability':								
σ_H	0.97		3.48		0.54		1.86	
$0,3*\sigma_H$	0.29		1.04		0.16		0.56	
x0-xa<0,3* σ_H ? (consequential instability)	NO		NO		NO		NO	
Test 'significant difference':								
t	2.21		0.28		1.29		0.09	
t-crit	2.23		2.23		2.23		2.23	
Significant difference	NO		NO		NO		NO	

Appendix 3: Copy of letter of invitation



HBM4EU: Announcement / invitation to participate in ICI / EQUAS study OPFR/Round 4

Title of ICI/EQUAS: OPFR in urine

Dear Colleagues,
within the frame of HBM4EU,

Prof. Dr. Jana Hajšlová & Dr. Dajana Dvoraková
University of Chemistry and Technology, Prague
Department of Food Analysis and Nutrition
Technická 3
166 28 Prague 6
Czech Republic

announce the 4th round of ICI/EQUAS for the determination of OPFR in urine. The aim of ICI/EQUAS exercises is to provide laboratories with an assessment of their analytical performance and reliability of their data in comparison with other laboratories and/or expert laboratories. This will aid in the quality improvement of analysis in human biomonitoring at each of the laboratories.

Participation is mandatory for laboratories analysing samples in the frame of HBM4EU.

Test samples

The matrix will be human urine. The participants will receive:

- 2 different materials of urine (1 sample of 5 ml each) for determination of OPFR in urine

Target biomarkers

Please see registration form for OPFR/Round 4 for the biomarkers potentially present in the test samples. We would be pleased if your laboratory could participate with the analysis of as most as possible OPFR.

LOQs should allow the analysis of OPFR in samples of the general population.

Calendar:

Deadline for registration	10-11-2019
Distribution of test samples (projected)	12-11-2019
Deadline for reporting the results (projected)	20-12-2019



Registration

For registration, please find attached a registration form for OPFR in serum. Please send it back to us by mail in case you want to register.

Upon registration, the participant will receive a lab-code to be used for submission of results.

Fee

For partners and linked-third parties of HBM4EU, participation is free of charge. Please note that the participant is responsible for custom clearance and associated costs if applicable.

Confidentiality:


All laboratory specific information will be treated confidentially, and will never be disclosed to third parties (government, accreditation bodies) except the HBM4EU QAU, without permission of the laboratory.

Contact information organiser:

Dr. Dajana Dvoraková
University of Chemistry and Technology, Prague
Department of Food Analysis and Nutrition
Technická 3
166 28 Prague 6
Czech Republic

Email of organiser: dajana.lankova@vscht.cz
Email for registration: VSCHT-hbm4EU@vscht.cz
Phone: 00420 22044 4312

Appendix 4: Copy of registration form for participation



HBM4EU: Registration form for participation in ICI / EQUAS study OPFR/Round 4.

Title of ICI/EQUAS: OPFR in urine

Please choose the OPFR you want to participate with.
We would appreciate your registration for as much biomarkers as possible.


Parameter	Participation	
	Yes	No
DPHP		
BCIPP		
BCEP		
BCIPP		

Participating laboratory:
name of institution

address of the laboratory

name of 1st contact person, telephone number and email address

name of 2nd contact person, telephone number and email address



Address for delivery of the test samples
name of institution

address of the laboratory

The above laboratory will participate in the ICI/EQUAS study **OPFR/Round 4**.
I agree with the conditions mentioned in the invitation letter, and that the laboratory will analyse the ICI/EQUAS samples using the same procedure as will be used for analysis of samples in the frame of HBM4EU, and submit results before the indicated deadline.

Name: _____ Signature _____

Date: _____

After signing this form, please scan and send the pdf to:
VSCHT-hbm4EU@vvscht.cz

Contact information organiser:
Dr. Dama Dvorakova
University of Chemistry and Technology, Prague
Department of Food Analysis and Nutrition
Technická 3
166 28 Prague 6
Czech Republic

Email: dama.lankova@vvscht.cz
Phone: 00420 22044 4312

Appendix 5: Copy of letter/instructions sent together with test samples



Please send the filled in result table together with this signed result submission form back to:

VSCHT-hbm4EU@vsccht.cz

If you have any questions or need any assistance, please contact:

Darina Dvorakova
Email: darina.dvorakova@vsccht.cz
Phone: 00420 22044 4312

Jana Hajšlova
Email: jana.hajslova@vsccht.cz
Phone 00420602833424

Contact information organiser:

Dr. Darina Dvorakova
University of Chemistry and Technology, Prague
Department of Food Analysis and Nutrition
Technická 3
166 28 Prague 6
Czech Republic

Prof. Ing. Jana Hajšlova, CSc.
(for the ICI/EQUAS organisers)



HBM4EU: Result submission form ICI/EQUAS study OPFR in urine Round 4.

Title of ICI/EQUAS: OPFR in urine

Laboratory name & code:

Contact person:

Institute:

Address:

Country:


For each of the analytes from this ICI/EQUAS round:

- provide the LOQ
- report NA for not analysed/not included in the scope of the method used
- report ND when not detected or detected below LOQ
- report a numerical value when found above LOQ, express to three significant figures (e.g. 0.543)

Date:

Signature:

Appendix 6: Copy of acknowledgement of receipt sent together with test samples



science and policy
for a healthy future


If you have any questions or need any assistance, please contact:

Darina Dvorakova
Email: darina.dvorakova@vscht.cz
Phone: 00420 22044 4312

Jana Hajsova
Email: jana.hajsova@vscht.cz
Phone 00420 602 833 424

Contact information organiser:
Dr. Darina Dvorakova
University of Chemistry and Technology, Prague
Department of Food Analysis and Nutrition
Technická 3
166 28 Prague 6
Czech Republic

Prof. Ing. Jana Hajsova, CSc.
(for the ICI/EQUAS organisers)



science and policy
for a healthy future

HBM4EU: Acknowledgement of receipt form ICI/EQUAS study OPFR/Round 4.

Title of ICI/EQUAS: OPFR in urine

Laboratory name:

Contact person:

Contents of parcel:
- 2 tubes with 5 ml urine

Please verify that the items listed below have been received and provide the information requested below:

Date of receipt (dd-mm-yyyy):

Code on tube	Damage/leakage	Remarks

Name: _____ **Signature:** _____

Date: _____

After signing this form, please scan and send the pdf. to: VSCHT-hbm4eu@vscht.cz

Appendix 7: Copy of method information form for participation in ICI/EQUAS



Method information form for participation in ICI/EQUAS OPFR in urine OPFR/Round 4.

Laboratory name & code	yes / no
ISO17025 accredited	
SAMPLE PREPARATION	
amount sample extracted (ml)	
Deconjugation	yes / no
chemical (reagent / pH / temp / time)	
enzymatic (enzyme / pH / temp / time)	
Extraction	
pH adjustment	
LLE (solvent(s) / time / shaking)	
SPE (material)	
Cleanup	
LLE (solvent(s))	
SPE (material)	
Derivatisation	
reagent	
INSTRUMENTAL ANALYSIS	
HPLC	
injection volume (µl)	
column stationary phase	
column L (mm) x ID (mm), dp (µm)	
temperature	
mobile phase A (specify the mobile additives)	
mobile phase B (specify the mobile additives)	
flow rate (ml/min)	
GC	
injector (splitless/PTV...)	
injection volume	
column stationary phase	
column L (mm) x ID (mm) dI (µm)	
carrier	
flow rate / inlet pressure	
Detection	
MS (single quadrupole quad/Q-Orbitrap/Q-TOF)	
MS manufacturer (system specification)	
Other	



QUANTIFICATION	
Use of internal standard (IS)	yes / no
isotopic label yes/no	
please specify the IS including purity used for each OPFRs	
DPHP	
BCEP	
BDCIPP	
BCIPP	
moment of addition (e.g. before deconjugation, to final extract...)	
response normalised to IS	yes / no
Calibration	
isotope dilution (addition to sample before extraction)	
isotope dilution (addition to final extract)	
standard addition (addition to sample before extraction)	
standard addition (addition to final extract)	
matrix-matched (addition to blank matrix before extraction)	
matrix-matched (addition to blank extract)	
solvent standards	
single level / multi level	yes / no
Correction for recovery	
Recovery value (%) (if applicable specify for each analyte)	
DPHP	
BCEP	
BDCIPP	
BCIPP	
Identification criteria used	
retention time tolerance (min or % deviation from reference standard)	
number of ions/transitions required for identification	
specify ions/transitions used for the quantification and confirmation	
DPHP - quantification	
DPHP - confirmation	
BCEP - quantification	
BCEP - confirmation	
BDCIPP - quantification	
BDCIPP - confirmation	
BCIPP - quantification	
BCIPP - confirmation	
ion ratio tolerance (% relative/absolute deviation from reference standard)	

Appendix 8: Graphical representation of the Z-scores

