



science and policy
for a healthy future

HBM4EU project

Determination of aflatoxin biomarkers
for acute and chronic exposure

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1. Overview

2. Aflatoxin biomarkers

3. Determination aflatoxin biomarkers

4. Quality Control and determination of “Unknowns”

- International Agency for Research on Cancer (IARC)
- Classification according to evidence of carcinogenicity to humans

Group	Classification	Mycotoxins
1	Carcinogenic to humans	aflatoxins
2A	Probably carcinogenic to humans	/
2B	Possibly carcinogenic to humans	ochratoxin A, sterigmatocystin and fumonisins
3	Not classifiable as to its carcinogenicity to humans	deoxynivalenol, nivalenol, T-2 toxin, diacetoxyscirpenol, zearalenone, citrinin and fusarenon-X
4	Probably not carcinogenic to humans	/

- As DON, AFB1 has validated biomarkers

- Acute exposure: AFB-N7-guanine

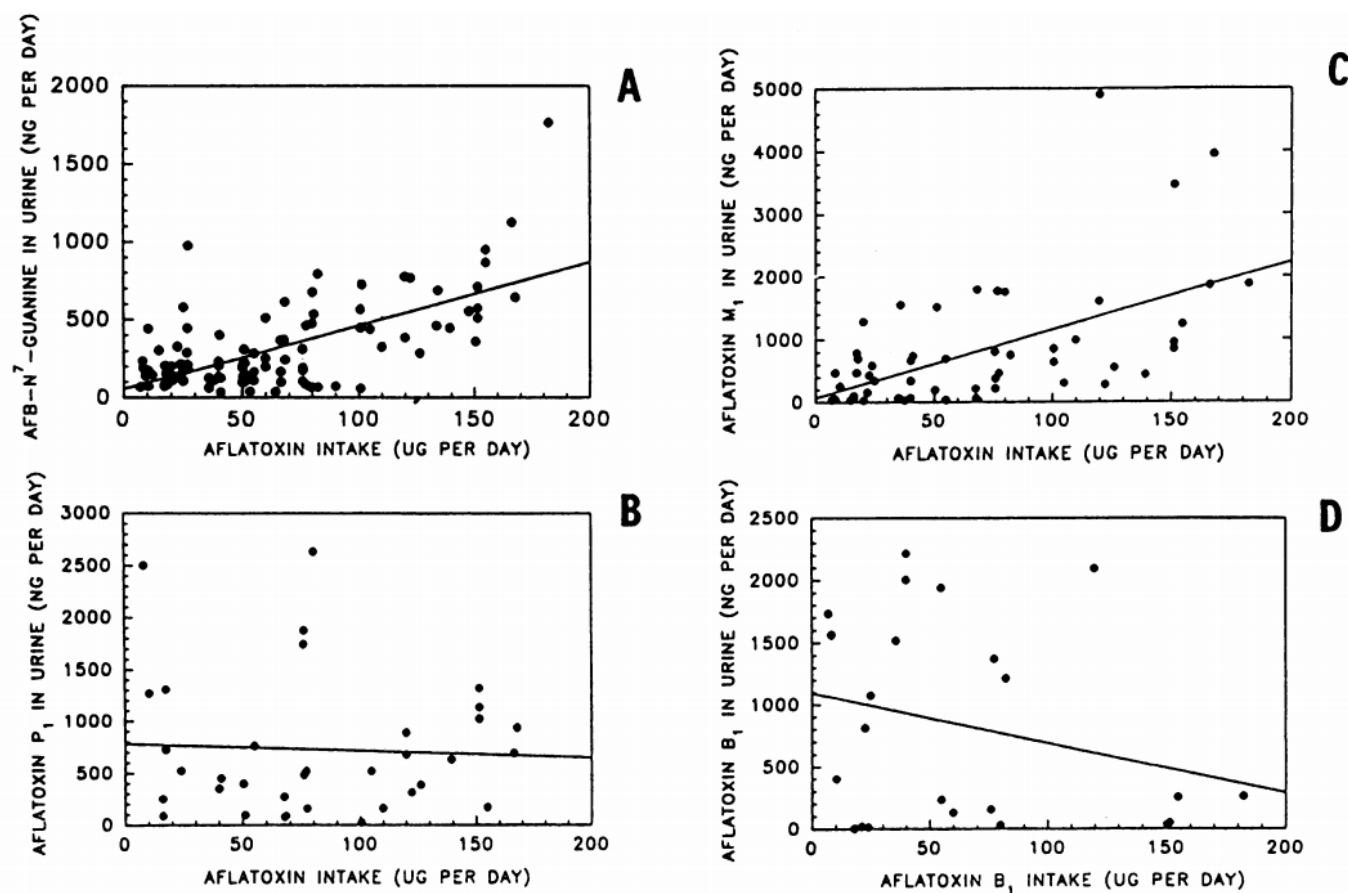


FIGURE 2. Linear regression analysis of daily (4) aflatoxin (AFB) N⁷-guanine, (B) AFM₁, (C) AFP₁, and (D) AFB₁ in urine compared with dietary aflatoxin intake from the previous day (5).

Groopman et al., 1992

- Acute exposure: AFB-N7-guanine
- Chronic exposure: AFB1-lysine
 - ✓ AFB1-lysine biomarker validated ELISA (Wild et al., 1992).
 - ✓ AFB1-lysine biomarker validated by LC-MS/MS (McMillan et al, 2018).



2.6 times more specific than ELISA technique

- Chronic exposure: AFB1-lysine

- ✓ AF-lysine biomarker validated ELISA (Wild et al., 1992).

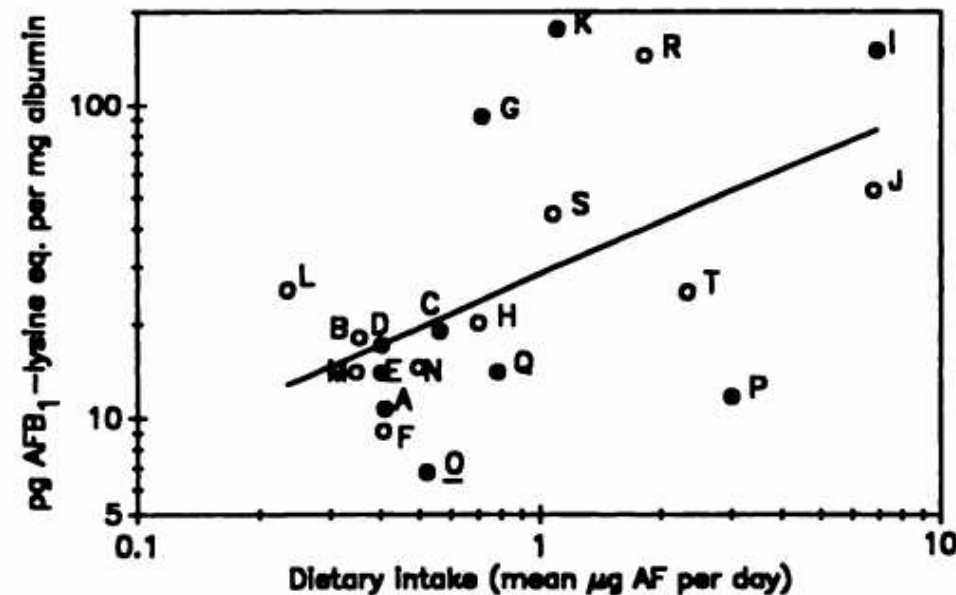
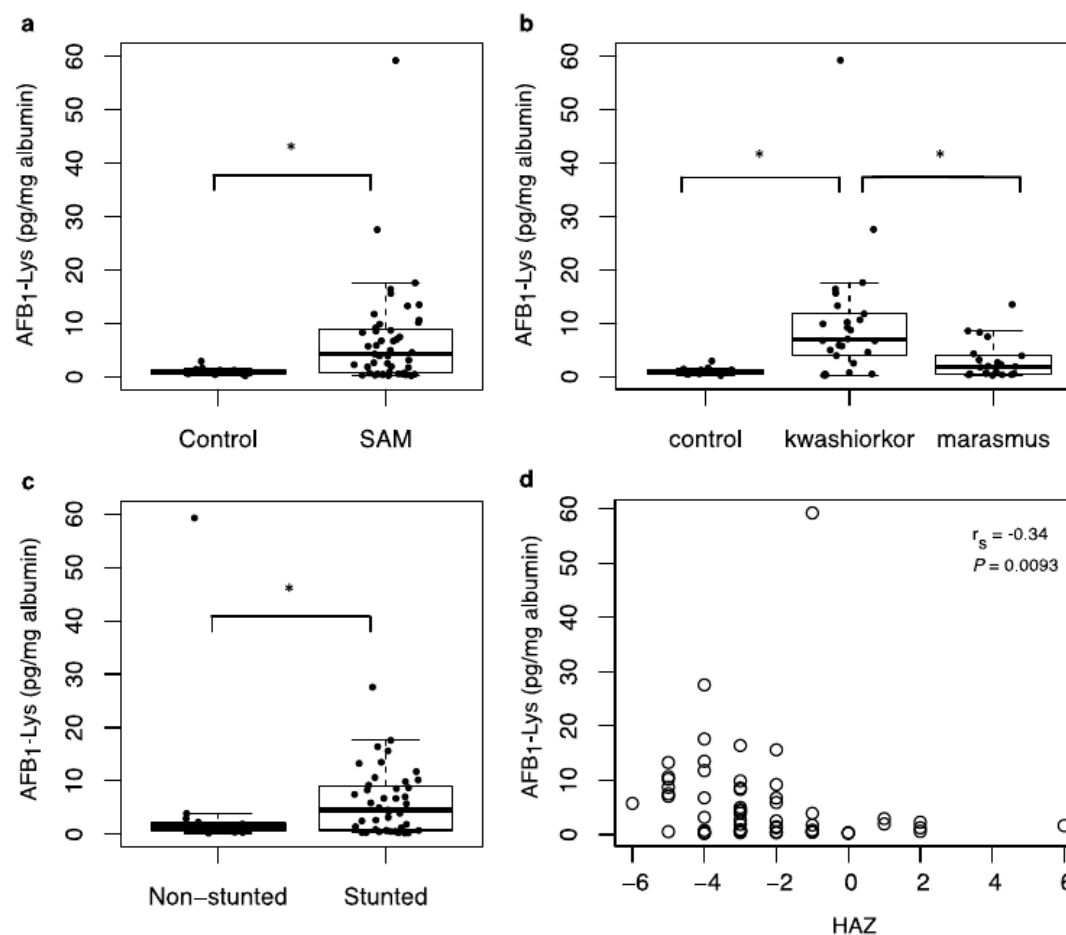


Fig. 4. Mean daily aflatoxin food intake over the 7-day period, plotted against the level of aflatoxin-albumin adduct on day 8 of the study. Each point represents one individual. ●, HBsAg carriers; ○, noncarriers. The letters next to the points represent the individuals in Table 1. Linear regression is plotted (correlation coefficient, $r = 0.55$; $P < 0.05$ on log-transformed values).

- Assess mycotoxin exposure with correct mycotoxin biomarker:



- However, AF biomarkers in urine:
 - 4 metabolic pathways:
 - ✓ O-dealkylation: AFP1
 - ✓ Keto-reduction: AFL
 - ✓ Epoxidation: AFB1-8,9-epoxide
 - ✓ Hydroxylation: AFM1, AFP1, AFQ1 or AFB2
 - **AFQ1>AFM1** (Mykkanen et al., 2005)
 - **AFP1>AFB1-N7-guanine** (Groopman et al., 1992).
 - Lack of commercial standards

- *In vitro* study

Liver microsomes + mycotoxin + Tris-HCl buffer

Phase I

Nicotinamide adenine dinucleotide
phosphate (NADPH)

Phase II

Uridine Diphosphate
Glucuronic Acid
(UDPGA)

Identification with High Resolution

- AFQ1
- AFP1



- *In vivo* study

Analysis of:

- AFB1 (standard)
- AFB2 (standard)
- AFG1 (standard)
- AFG2 (standard)
- AFM1 (standard)
- AFB1-lys (synthesised)
- AFB1-N7-guanine (synthesised)
- AFQ1 (no standard)
- AFP1 (no standard)
- Isotolabelled C13 AFB1 (Internal standard)

- *In vivo* study
 - Extraction method:
 - IAC columns:
 - ✓ NOT highly checked for AF conjugates.
 - ✓ Expensive.
 - ✓ Long time.
 - ELISA:
 - ✓ Higher LOD.
 - ✓ Less specific.
 - Dilute and shoot:
 - ✓ Small volume.
 - ✓ Fast.
 - Liquid/Liquid extraction:
 - ✓ You can concentrate.
 - ✓ Check recovery.

- In vivo* study

Country (matrix)	Mycotoxins	Extraction method	Limit of detection (ng/mL)	Average (ng/mL)	Reference
Belgium (Urine)	AFB1 AFB2 AFG1 AFG2 AFM1	Liquid/Liquid	0.001	Not detected	Heyndrickx et al., 2015
Belgium (Urine)	AFM1 AF-guanine	Liquid/Liquid with SPE column	0.01 0.85	Not detected	Njumbe Ediage et al., 2012
Italy (Urine)	AFB1 AFB2 AFG1 AFG2 AFM1	IAC Column	0.010 0.006 0.006 0.004 0.002	0.010 (0.8 %) 0.007 (0.8%) 0.058 (0.8%) 0.057 (11.1%) 0.042 (73.7%)	Ferri et al., 2017
Italy (Plasma)	AFB1 AFB2 AFG1 AFG2 AFM1	IAC Column	0.025 0.025 0.006 0.006 0.025	Not detected	Ferri et al., 2017

- In vivo* study

Coutnry (matrix)	Mycoto xins	Extraction method	Limit of detection (ng/mL)	Average (ng/mL)	Reference
Ethiopia (Urine)	AFB1 AFB2 AFG1 AFG2 AFM1	Dilute and shoot	0.025	- 0.047 (4.5%) 0.061 (2.5%) 0.068 (3%) 0.064 (7%)	Ayelign et al., 2017
Cameroon (Urine)	AFM1 AFN7guanine	Liquid/liquid	0.01 0.83	0.33 (max = 4.7) (14 %)	Njumbe Ediage et al., 2013
Nigeria (Urine)	AFM1	ELISA	0.06	0.27 (98.8 %)	Chibundu et al., 2018
Malawi (Plasma)	AFB1lys	Liquid/Liquid	0.002	0.023 (73%)	Seetha et al., 2018
Nigeria (Plasma)	AFB1lys	Liquid/Liquid	0.022	0.0026	McMillan et al., 2018

QA-CONTROL in BIOMARKER ANALYSIS using LC-MS/MS

- First line control
- Second line control
- Third line control
- Identification of 'unknowns'

- First line control
 - ✓ Assurance of a good performance of the analytical device and the correctness of the acquired results.
 - ✓ Analysis according to a quality control (QC)-scheme of a serie of unknown samples

- First line control
 1. A mix of calibrants in pure solvent = standard mix.
 2. Sample with pure injection solvent = mobile phase.
 3. Blank sample (urine/plasma/...).
 4. Spiked samples for the calibration curve (min. 5 points).
 5. Sample with pure injection solvent = mobile phase.
 6. Ten unknown samples.
 7. Control spike.
 8. Ten unknown samples.
 9. Control spike.

- **First line control**

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8. Ten unknown samples.
- 9. Control spike.**

What	Why	QC criteria	Consequences if QC-criteria are not fulfilled								
Control spike	Check quantification during injection-sequence	- The recovery of each compound needs to range between (concentration dependent):<table><tr><th>Concentratic</th><th>Interval</th></tr><tr><td>≤ 1 µg/kg</td><td>50% – 120%</td></tr><tr><td>> 1 µg/kg – 10 µg/kg</td><td>70% - 110%</td></tr><tr><td>>10 µg/kg</td><td>80% - 110%</td></tr></table> - Or as determined as in the method validation (compound specific) - Set-up a trend analysis!	Concentratic	Interval	≤ 1 µg/kg	50% – 120%	> 1 µg/kg – 10 µg/kg	70% - 110%	>10 µg/kg	80% - 110%	- When recovery is NOT OK: quantification of all samples between control spike and previous control spike are not reliable. Re-analysis! - The recovery of the control spike needs to be followed-up over a longer period. Visible trends need to be investigated when they falls out of an interval.
Concentratic	Interval										
≤ 1 µg/kg	50% – 120%										
> 1 µg/kg – 10 µg/kg	70% - 110%										
>10 µg/kg	80% - 110%										

- Second line control
 - ✓ Periodical evaluation (eg 2/year).
 - ✓ To check method with the acquired method validation (accuracy, LOD/LOQ, ...).
 - ✓ New analyst.
 - ✓ ...

 - ✓ Analysis of certified reference material.
 - ✓ Analysis of spiked sample by a third person.
 - ✓ Analysis of a blind, duplicated sample.

- Third line control
 - ✓ Quality control organised by an independent external organisation.
 - ✓ Interlaboratory test.
 - ✓ To compare and evaluate the performance of your developed method with other methods.
 - ✓ At least 1 x 3 years.

- Identification of unknowns
- Fulfilment of **4 identification criteria**:
 1. **Minimum of 3** to more identification points because
2 **MRM-transitions** are present.
 2. **S/N** every MRM-transition > 3 .
 3. **Relative peak-area** of selected ions has to correspond with those ions of the spike with a comparable concentration in an acceptable deviation.
 4. **Relative retention time** of each MRM-transition should be within a range of 2.5% of the relative retention time of the spiked sample with a comparable concentration.

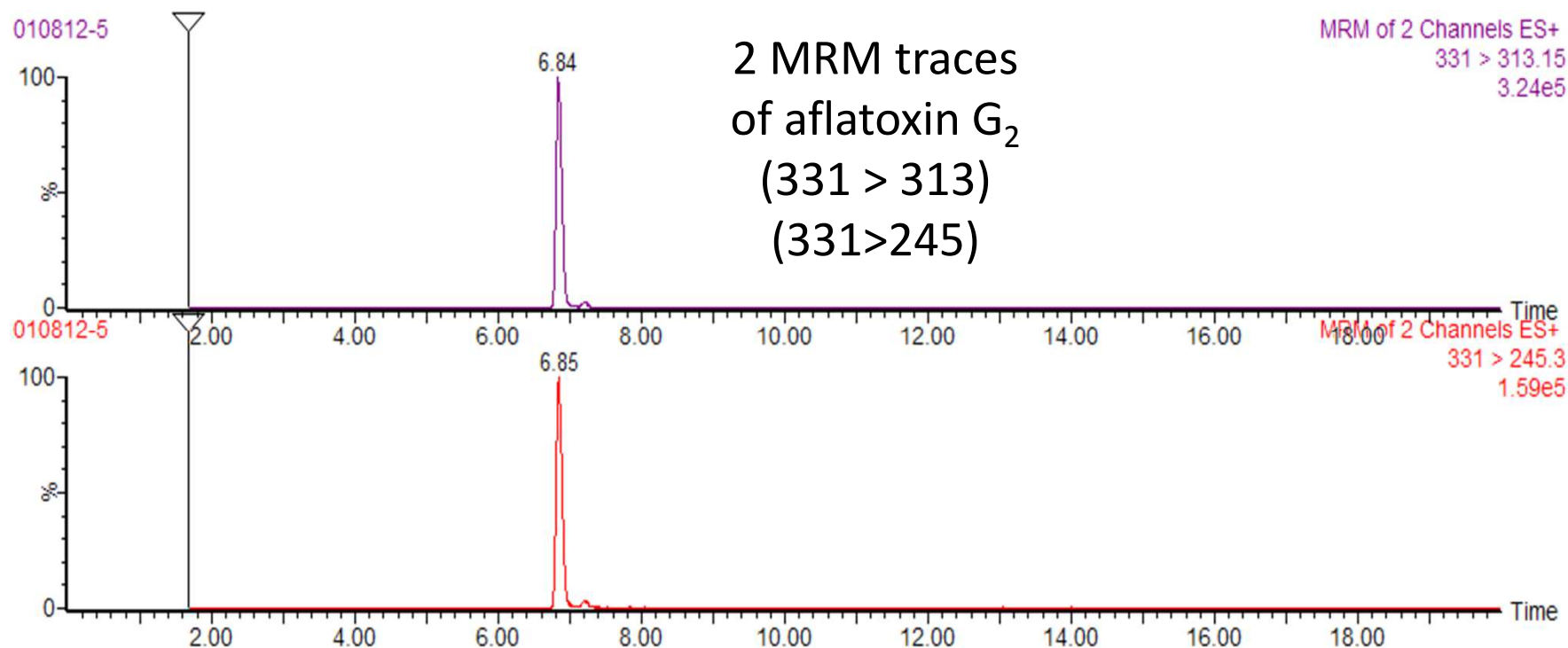
- Identification of unknowns
- Fulfilment of **4 identification criteria**:
 - 1. Minimum of 3** to more identification points because
 - 2 MRM-transitions** are present.

The relationship between a range of classes of mass fragment and identification points earned

MS technique	Identification points earned per ion
Low resolution mass spectrometry (LR)	1,0
LR-MS ⁿ precursor ion	1,0
LR-MS ⁿ transition products	1,5
HRMS	2,0
HR- MS ⁿ precursor ion	2,0
HR-MS ⁿ transition products	2,5

X 2 (MRM transitions) = 3 identification points

- Identification of unknowns
- Fulfilment of **4 identification criteria:**
 - 2. S/N** every MRM-transition > 3.

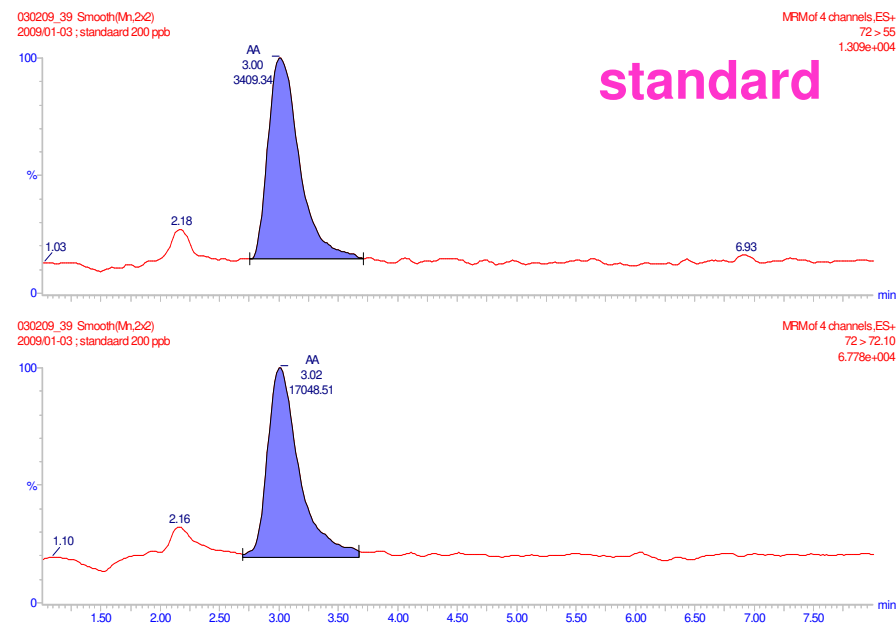
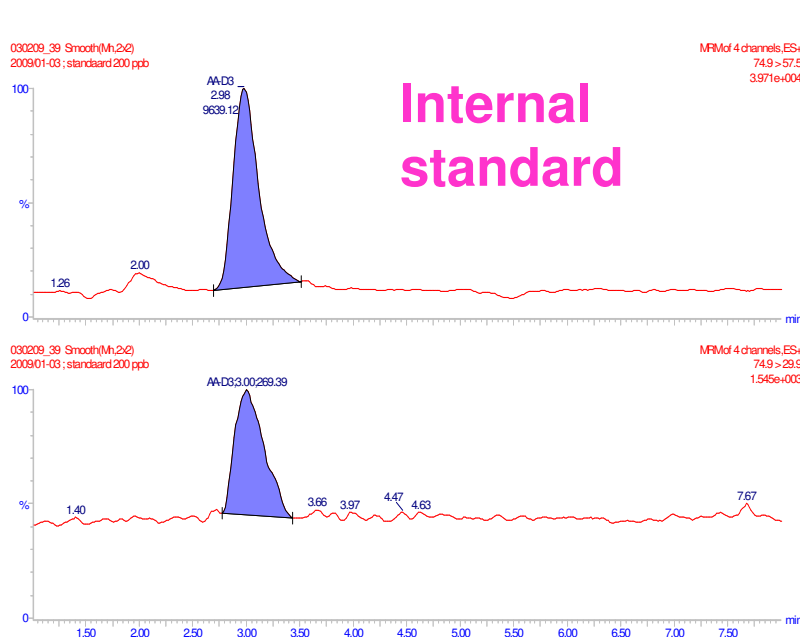


- Identification of unknowns
- Fulfilment of **4 identification criteria**:
 - 3. Relative peak-area** of selected ions has to correspond with those ions of the spike with a comparable concentration in an acceptable deviation.

<i>Relative intensity (% of mean peak)</i>	<i>Accepted limits LC-MS/MS</i>
> 50%	± 20%
> 20% – 50%	± 25%
> 10% - 20%	± 30%
≤ 10%	± 50%

- Identification of unknowns
- Fulfilment of **4 identification criteria:**
 - 3. Relative peak-area** of selected ions has to correspond with those ions of the spike with a comparable concentration in an acceptable deviation.
 - Compare with spike with similar concentration.
 - Relative peak area spike: $(\text{area ion } 331>313)/(\text{area ion } 331>245) = x$
 - Range determined on spike: $[x - \text{limit}; x + \text{limit}]$.
 - Relative peak area unknown: $(\text{area ion } 331>313)/(\text{area ion } 331>245) = z$
 - $x - \text{limit} < z < x + \text{limit}$.

- Identification of unknowns
- Fulfilment of **4 identification criteria**:
 - 4. Relative retention time** of each MRM-transition should be within a range of 2.5% of the relative retention time of the spiked sample with a comparable concentration.



- Identification of unknowns
- Fulfilment of **4 identification criteria:**
 - 4. Relative retention time** of each MRM-transition should be within a range of 2.5% of the relative retention time of the spiked sample with a comparable concentration.
 - Compare with calibration standard (spike)
 - Relative retention time spike: $(RT\ MYCO)/(RT\ IS) = r$
 - Range calculated on spike: $[r - 2.5\%; r + 2.5\%]$
 - Relative retention time unknown: $(RT\ Unknown)/(RT\ IS) = t$
 - $r - 2.5\% < t < r + 2.5\%$

- Identification of unknowns

- Fulfilment of 4 identification criteria

1. minimum of 3 to more identified MRM-transitions are present because 2
2. S/N every MRM-transition
3. relative peak-area to correspond with those of the reference compound a comparable concentration
4. relative retention time of each MRM-transition should be close to the relative retention time of the reference compound with a comparable concentration.

Only when these 4 criteria are fulfilled, you can state that the mycotoxin biomarker is PRESENT in your unknown sample!

The logo consists of the word "MYTOX" in a bold, sans-serif font. "MY" is light blue, "TO" is dark blue, and "X" is dark blue. The letter "O" is replaced by a stylized white icon of a cell or microorganism with three dots inside a circle.The logo consists of the word "MYTOX" in a bold, sans-serif font. "MY" is orange, "TO" is dark red, and "X" is dark red. The letter "O" is replaced by a stylized white icon of a cell or microorganism with three dots inside a circle. Below "MYTOX" is the word "SOUTH" in a smaller, dark red, sans-serif font.

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